## This Page Is Inserted by IFW Operations and is not a part of the Official Record

## **BEST AVAILABLE IMAGES**

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images may include (but are not limited to):

- BLACK BORDERS
- TEXT CUT OFF AT TOP, BOTTOM OR SIDES
- FADED TEXT
- ILLEGIBLE TEXT
- SKEWED/SLANTED IMAGES
- COLORED PHOTOS
- BLACK OR VERY BLACK AND WHITE DARK PHOTOS
- GRAY SCALE DOCUMENTS

### IMAGES ARE BEST AVAILABLE COPY.

As rescanning documents will not correct images, please do not report the images to the Image Problem Mailbox.

## {Exhibit 39}

Rosswell, D.F. and White E.H., "The Chemiluminescence of Luminol and Related Hydrazides" in Methods in Enzymol. Vol LV II, Bioluminescence and Chemiluminescence, DeLuca, M.A. (Ed.) 409-423 (1978)

Methods in Enzymology

Volume LVII

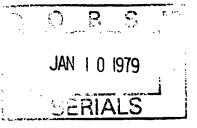
# Bioluminescence and Chemiluminescence

**EDITED BY** 

Marlene A. DeLuca

DEPARTMENT OF CHEMISTRY

UNIVERSITY OF CALIFORNIA AT SAN DIEGO
LA JOLLA, CALIFORNIA





ACADEMIC PRESS New York San Francisco London 1978
A Subsidiary of Harcourt Brace Jovanovich, Publishers

Ref. 9 601 .C733 V.57 C.1

COPYRIGHT © 1978, BY ACADEMIC PRESS, INC. ALL RIGHTS RESERVED.

NO PART OF THIS PUBLICATION MAY BE REPRODUCED OR TRANSMITTED IN ANY FORM OR BY ANY MEANS. ELECTRONIC OR MECHANICAL, INCLUDING PHOTOCOPY, RECORDING. OR ANY INFORMATION STORAGE AND RETRIEVAL SYSTEM. WITHOUT PERMISSION IN WRITING FROM THE PUBLISHER.

ACADEMIC PRESS, INC.
111 Fifth Avenue, New York, New York 10003

United Kingdom Edition published by ACADEMIC PRESS, INC. (LONDON) LTD. 24/28 Oval Road, London NW1 7DX

Library of Congress Cataloging in Publication Data Main entry under title:

Bioluminescence and chemiluminescence.

(Methods in enzymology; v. 57)
Includes bibliographical references and index.

1. Bioluminescence. 2. Chemiluminescence.
I. DeLuca, Marlene. II. Title.

QP601.M49 vol. 57 [QH641] 574.1'925'08s

[574.1'9125]
ISBN 0-12-181957-4 78-13442

PRINTED IN THE UNITED STATES OF AMERICA

#### Table of Contents

CONTRIBUTORS TO VOLUME LVII		ix
Preface		xiii
Volumes in Series		xv
Section I. Firefly Luc	ciferase	
1. Purification and Properties of Firefly Luciferase	MARLENE DELUCA AND W. D. McElroy	3
2. Synthesis of Firefly Luciferin and Structural Analogs	LEMUEL J. BOWIE	15
3. Preparation of Partially Purified Firefly Luciferase Suitable for Coupled Assays	HANS N. RASMUSSEN	28
4. Firefly Luciferase Assay for ATP Production by Mitochondria	JOHN J. LEMASTERS AND CHARLES R. HACKENBROCK	36
5. Measurement of Photophosphorylation and ATP- ase Using Purified Firefly Luciferase	ARNE LUNDIN AND MARGARETA BALTSCHEFFSKY	50
<ol> <li>Determination of Creatine Kinase Isoenzymes in Human Serum by an Immunological Method Using Purified Firefly Luciferase</li> </ol>	Arne Lundin	56
7. Determination of Bacterial Content in Fluids	EMMETT W. CHAPPELLE, GRACE L. PICCIOLO, AND JODY W. DEMING	6.
8. Biomass and Adenylate Energy Charge Determina- tion in Microbial Cell Extracts and Environmen- tal Samples	Osmund Holm-Hansen and David M. Karl	7:
<ol><li>Determination of GTP, GDP, and GMP in Cell and Tissue Extracts</li></ol>	DAVID M. KARL	8:
<ol> <li>Measurement of the Activity of Cyclic Nucleotide Phosphodiesterases with Firefly Luciferin- Luciferase Coupled Assay Systems</li> </ol>	RICHARD FERTEL AND BENJAMIN WEISS	9
<ol> <li>Rapid Microassay for the Calcium-Dependent Pro- tein Modulator of Cyclic Nucleotide Phospho- diesterase</li> </ol>	JOHN C. MATTHEWS AND MILTON J. CORMIER	10
12. ATP-Labeled Ligands and Firefly Luciferase for Monitoring Specific Protein-Binding Reactions	ROBERT J. CARRICO, R. DON JOHNSON, AND ROBERT C. BOGUSLASKI	11

#### Section II. Bacterial Luciferase 13. Bacterial Bioluminescence: An Overview J. W. HASTINGS 125 14. Bacterial Luciferase: Assay, Purification, and J. W. HASTINGS, THOMAS O. BALDWIN, AND MIRIAM Z. Properties NICOLI 135 15. Isolation, Identification, and Manipulation of Luminous Bacteria KENNETH H. NEALSON 153 16. Isolation and Characterization of Luminescence System Mutants in Bacteria THOMAS W. CLINE 166 17. Preparation of the Subunits of Bacterial SHIAO-CHUN TU Luciferase 171 18. Preparation of Luciferases Containing Chemically Modified Subunits EDWARD A. MEIGHEN 174 19. Quantitation of Malate, Oxaloacetate, and Malate Dehydrogenase PHILIP E. STANLEY 181 20. Bioassay for Myristic Acid and Long-Chain Al- S. ULITZUR AND J. W. dehydes HASTINGS 189 21. The Luciferase Reduced Flavin-Hydroperoxide J. W. HASTINGS AND JAMES E. BECVAR Intermediate 194 22. Bacterial Luciferase as a Generalized Substrate for the Assay of Proteases THOMAS O. BALDWIN 198 23. Immobilization of Bacterial Luciferase and Oxidoreductase and Assays Using Immobilized EDWARD JABLONSKI AND Enzymes MARLENE DELUCA 202 24. Quantitation of Picomole Amounts of NADH, NADPH, and FMN Using Bacterial Luciferase Philip E. Stanley 215 25. Luminous Bacteria as an Oxygen Indicator B. CHANCE AND R. OSHINO 223 26. Purification of a Blue-Fluorescent Protein from the Bioluminescent Bacterium Photobacterium JOHN LEE AND PRASAD phosphoreum KOKA 226 Section III. Renilla reniformis Luciferase 27. Applications of Renilla Bioluminescence: An Introduction 237 MILTON J. CORMIER 28. A Bioluminescence Assay for PAP (3'., 5'-Di- James Michael Anderson, phosphoadenosine) and PAPS (3'-Phospho- Kazuo Hori, and adenylyl Sulfate) MILTON J. CORMIER 244 29. Protein-Protein Interactions as Measured by WILLIAM W. WARD AND Bioluminescence Energy Transfer in Renilla 257 MILTON J. CORMIER

The state of the s

	Section IV. Aequ	orin	
30.	Introduction to the Bioluminescence of Medusae, with Special Reference to the Photoprotein Aequorin	Frank H. Johnson and Osamu Shimomura	271
31.	Practical Aspects of the Use of Aequorin as a Calcium Indicator: Assay, Preparation, Microinjection, and Interpretation of Signals	JOHN R. BLINKS, PATRICK H. MATTINGLY, BRIAN R. JEWELL, MENNO VAN LEEUWEN, GARY C. HARRER, AND DAVID G. ALLEN	292
	Section V. Cypric	lina	•
32.	Introduction to the Cypridina System	Frank H. Johnson and Osamu Shimomura	331
33.	Cypridina Luciferin and Luciferase	FREDERICK I. TSUJI	364
	Section VI. Earthworm Bio	oluminescence	
34.	Assaying Hydrogen Peroxide Using the Earthworm Bioluminescence System	MICHAEL G. MULKERRIN AND JOHN E. WAMPLER	375
	Section VII. Pholas of	dactylus	
35	Purification and Properties of <i>Pholas dactylus</i> Luciferin and Luciferase	A. M. MICHELSON	385
	Section VIII. Chemilumines	cent Techniques	,
36	. The Chemiluminescence of Luminol and Related Hydrazides	DAVID F. ROSWELL AND EMIL H. WHITE	409 🗸
37	. Monitoring Specific Protein-Binding Reactions with Chemiluminescence	HARTMUT R. SCHROEDER, ROBERT C. BOGUSLASKI, ROBERT J. CARRICO, AND ROBERT T. BUCKLER	424
38	Chemiluminescence Detection of Enzymically Generated Peroxide	W. RUDOLF SEITZ	445
39	P. The Generation of Chemiluminescence (CL) by Phagocytic Cells	MICHAEL A. TRUSH, MARK E. WILSON, AND KNOX VAN DYKE	462

viii	TABLE OF CONTENTS

40. Chemiluminescence from Electron-Transfer Processes	LARRY R. FAULKNER	494
Section IX. Instrumentation	n and Methods	
41. Construction of Instrumentation for Bioluminescence and Chemiluminescence Assays	James Michael Anderson, George J. Faini, and John E. Wampler	529
42. The Simultaneous Determination of Heat Changes and Light Production	S Neal Langerman	540
43. Characteristics of Commercial Instruments and Reagents for Luminescent Assays	GRACE LEE PICCIOLO, JODY W. DEMING, DAVID A. NIBLEY, AND EMMETT W. CHAPPELLE	550
44. Excited States and Absolute Calibrations in Bioluminescence	n H. H. Seliger	560
AUTHOR INDEX		601
SUBJECT INDEX		616

FILE COPY

## [36] The Chemiluminescence of Luminol and Related Hydrazides

By DAVID F. ROSWELL and EMIL H. WHITE

Chemiluminescence is the production of light by chemical reactions. Electronically excited states are formed in the chemical reaction, and light emission from these completes the process. The relationship between chemiluminescence and absorption/emission phenomena, such as fluorescence, are shown in Fig. 1.

A surprisingly large number of chemical reactions are chemiluminescent. <sup>1-4</sup> This paper, however, will deal only with a rather famous example, the chemiluminescence of luminol [5-amino-2,3-dihydrophthalazine-1,4-dione (compound I)]; some derivatives and analogs will also be discussed.

The first report of chemiluminescence from luminol was made by Albrecht<sup>5</sup> in 1928. Since that time the reaction of luminol and other derivatives of the general hydrazide structure<sup>6</sup> have been studied extensively. A number of reviews on this subject have appeared over the last 15 years, 7-10 and hence the historical aspects of the problem will be treated only briefly.

- <sup>1</sup> K.-D. Gundermann, Top. Curr. Chem. 46, 61 (1974).
- <sup>2</sup> E. H. White and D. F. Roswell, Acc. Chem. Res. 3, 54 (1970).
- <sup>3</sup> M. J. Cormier, D. M. Hercules, and J. Lee, eds., "Chemiluminescence and Bioluminescence." Plenum, New York, 1973.
- <sup>4</sup> K.-D. Gundermann, "Chemilumineszenz Organischer Verbindungen." Springer-Verlag, Berlin and New York, 1968.
- <sup>5</sup> H. O. Albrecht, Z. Phys. Chem. 136, 321 (1928).
- 6 Luminol as well as being named as a phthalazinedione is also referred to as 3-aminophthalic hydrazide.
- <sup>7</sup> E. H. White, in "Light and Life" (W. D. McElroy and B. Glass, eds.), 1st ed., p. 183. Johns Hopkins Press, Baltimore, Maryland, 1961.
- <sup>8</sup> K.-D. Gundermann, Angew. Chem., Int. Ed. 4, 566 (1965).
- <sup>9</sup> F. McCapra, Quart. Rev. Chem. Soc., 20, 485 (1966).
- <sup>10</sup> J. W. Haas, J. Chem. Educ. 44, 396 (1967).

METHODS IN ENZYMOLOGY, VOL. LVII

Copyright © 1978 by Academic Press, Inc. All rights of reproduction in any form reserved. ISBN 0-12-181957-4

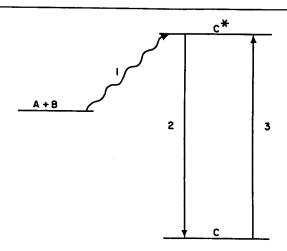


Fig. 1. Chemiluminescence, path 1 + 2; fluorescence, path 2; absorption, path 3.

#### **Reaction Conditions**

The chemiluminescent reactions of luminol are all oxidations [Eq. (1)]. A wide variety of reaction conditions have been employed; the reaction can be carried out either in protic media or in aprotic solvents such as dimethyl sulfoxide (DMSO), hexamethylphosphoric acid triamide (HMPT), or dimethylformamide (DMF). In aprotic media, only oxygen and a strong base are required for chemiluminescence.<sup>7</sup>

Water is the most common protic solvent, although the reaction has been studied in the lower alcohols as well. The reaction in these solvents usually requires base, an oxidizing agent, and either oxygen or a peroxide, depending on the system. Hypochlorite, ferricyanide, and persulfate are commonly used oxidizing agents. In place of the oxidizing agents, chelated transition metal complexes, pulse radiolysis, 11 or sonic waves 12 have been used in the presence of either oxygen or peroxides.

<sup>11</sup> J. H. Baxendale, Chem. Commun. 1971, 1490 (1971).

<sup>12</sup> E. N. Harvey, J. Am. Chem. Soc. 61, 2392 (1939).

One of the simplest systems involves adding luminol to a solution of hydrogen peroxide in dilute sodium carbonate that has been partially neutralized immediately before the luminol is added.<sup>13</sup>

#### Structural Effects

Early workers investigated the effect of gross structural modifications of luminol [Eq. (1)]. <sup>14-16</sup> Compounds isomeric to (I) in the heterocyclic ring—compounds (II) and (III), as well as N or O methylation products (IV and V, respectively)—were nonchemiluminescent.

Analogs in which the amino group was replaced by other substituents, or analogs with substitution anywhere in the nonheterocyclic ring were generally found to be chemiluminescent. These compounds vary in their light-producing ability; e.g., compounds (VI)¹ and (VII)¹³ are more chemiluminescent than luminol whereas compounds (VIII) and (IX) are of lower efficiency.¹⁴

$$(CH_{3}CH_{2})_{2}N$$

$$(VII)$$

$$(VIII)$$

$$(VIII)$$

$$(VIII)$$

$$(IX)$$

- <sup>13</sup> E. H. White and R. B. Brundrett, in "Chemiluminescence and Bioluminescence" (M. J. Cormier, D. M. Hercules, and J. Lee, eds.), p. 231. Plenum, New York, 1973.
- <sup>14</sup> H. D. K. Drew and F. H. Pearman, J. Chem. Soc. London 1937, 586 (1937).
- <sup>15</sup> H. D. K. Drew and R. F. Garwood, J. Chem. Soc. London 1939, 836 (1939).
- <sup>16</sup> E. H. Huntress and J. V. K. Gladding, J. Am. Chem. Soc. 64, 2644 (1942).

THE REPORT OF THE PARTY OF THE

The chemiluminescence quantum yield  $(\Phi_{ch})$ , where  $\Phi_{ch}=$  photons produced/molecules reacted, is used as the measure of light-producing ability. For luminol,  $\Phi_{ch}=0.01$ ; and for compound (VII),  $\Phi_{ch}=0.08$ . This quantum yield may also be defined as  $\Phi_{ch}=\Phi_R\Phi_{ES}\Phi_{Fl}$ , where  $\Phi_R$  is the fraction of reacting molecules following the correct chemical path,  $\Phi_{ES}$  is the fraction of excited states produced, and  $\Phi_{Fl}$  is the fluorescent quantum yield of the emitter. The light-producing ability of luminol analogs is thus a function of three factors; of these,  $\Phi_{ES}$  and  $\Phi_{Fl}$  appear to be the most important.<sup>17,18</sup>

#### **Products**

Under the mild reaction conditions possible in aprotic solvents, the organic product of the luminol oxidation is 3-aminophthalate (X), which is

produced almost quantitatively. <sup>19</sup> Nitrogen is evolved and oxygen is consumed in essentially molar amounts. <sup>19</sup> In protic media the reaction conditions are so vigorous that degradation of the initially formed aminophthalate occurs, but the overall reaction is believed to be the same.

#### The Light Emitter

In both protic and aprotic media the product phthalate ion, (X), is found in an electronically excited state and is responsible for the observed emission. The identity of the fluorescence spectrum of the product with the chemiluminescence emission spectra of the reaction (see Fig. 2) establishes this point. As shown in Fig. 2a, the emission from the oxidation of luminol, (I), in aqueous media occurs at 425 nm and is superimposable on the fluorescence spectra of the total reaction products and also on the spectrum of 3-aminophthalate ion. In aprotic media the emission maxima are shifted to longer wavelengths. The long-wavelength emission has been attributed to the quininoid tautomer, (XI), of (X). This species would not be present in

<sup>&</sup>lt;sup>17</sup> R. B. Brundrett, D. F. Roswell, and E. H. White, J. Am. Chem. Soc. 94, 7536 (1972).

<sup>&</sup>lt;sup>18</sup> R. B. Brundrett and E. H. White, J. Am. Chem. Soc., 96, 7497 (1974).

<sup>&</sup>lt;sup>19</sup> E. H. White, O. C. Zafiriou, H. M. Kagi, and J. H. M. Hill, J. Am. Chem. Soc. 86, 940 (1964).

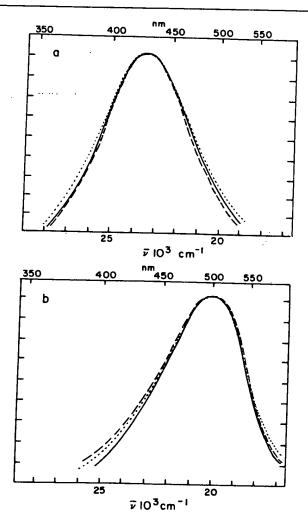


Fig. 2. Fluorescence and chemiluminescence emission spectra: (a) solvent is water; (b) dimethyl sulfoxide. ———, chemiluminescence of luminol; . . . . . , fluorescence of 3-aminophthalate; ----, fluorescence of total reaction mixture. Permission to reprint is granted by the American Chemical Society. J. Am. Chem. Soc. 95, 2610 (1973).

highly aqueous media<sup>20</sup> because of the lowered basicity of a carboxylate group hydrogen-bonded to water molecules. In mixed solvent systems (DMSO-H<sub>2</sub>O) both emissions occur, the 425-nm band increasing with water concentration. These reaction systems show a one-to-one correlation between chemiluminescence and the fluorescence of (X) as long as the base used is a quaternary ammonium hydroxide.

If other bases are used, this correspondence is lost; in a system of 90% dimethyl sulfoxide (DMSO)-water made basic with alkali metal hydroxides (MOH), the shorter-wavelength emission band is more pronounced in 3-aminophthalate fluorescence than in luminol chemiluminescence when  $M = Na^+$  or  $K^{+20}$  (Fig. 3).

The fluorescence spectrum of 3-aminophthalate shows a marked increase in the shorter-wavelength emission band with increasing sodium ion concentration, and at high sodium ion concentrations the longer-wavelength band is almost lost. The absorption spectrum of 3-aminophthalate ion shifts with increasing sodium ion concentration, suggesting that there is considerable ion paring in the ground state. Because of the short lifetime of the excited state, this ion pairing persists in the excited state. The ion pair of aminophthalate ion, (X), would be expected to have a lower electron density at the carbonyl moiety of the carboxylate group, and thus be less able to bring about the intramolecular proton transfer required to form the quininoid form (XI), which is responsible for the long-wavelength emission. Thus, metal ions form ion pairs with (X), which fluoresce at about 425 nm, whereas the free ion can pass over to the quininoid form, (XI), which fluoresces at about 510 nm.

The differences in the fluorescence emission spectra of 3-aminophthalate ion and the chemiluminescence of luminol (Fig. 3) can also be explained by the ion-pair effect. The fraction of excited aminophthalate ions in the ion-pair form differs in fluorescence and chemiluminescence. The fraction in fluorescence is essentially determined by the ground-state equilibrium. The fraction of ion pairs in chemiluminescence is determined

E. H. White, D. F. Roswell, C. C. Wei, and P. D. Wildes, J. Am. Chem. Soc. 94, 6223 (1972); see also J. Lee and H. H. Seliger, Photochem. Photobiol. 15, 227 (1972) and J. D. Gorsuch and D. M. Hercules, Photochem. Photobiol. 15, 567 (1972).

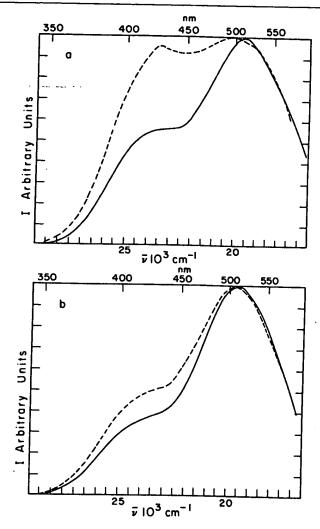


Fig. 3. Luminol chemiluminescence (———) and 3-aminophthalate fluorescence (---) in 30 mol % water, 70 mol % dimethyl sulfoxide with (a) 20 mM sodium hydroxide; (b) 20 mM potassium hydroxide. Permission to reprint is granted by the American Chemical Society. J. Am. Chem. Soc. 95, 2610 (1973).

at the transition state of the reaction, and in the present case this value is lower than that in fluorescence. It should be pointed out that the above observations pertain to luminol and that usually the excited states formed in chemiluminescence and fluorescence are the same.

The general picture of the chemiluminescence of cyclic hydrazides—that light emission occurs from the corresponding phthalate ion [Eq. (2)]—appears to hold with only rare exceptions. The parent compound (R=H,

phthalic hydrazide) is essentially nonchemiluminescent in the protic system;<sup>21</sup> this is not surprising, since phthalate ion under the reaction conditions is not fluorescent. In aprotic solvents such as DMSO, however, the reaction of phthalic hydrazide with base and oxygen does give a yellow emission of moderate intensity, yet under the same conditions phthalate ion is again not fluorescent. The emission spectrum of this chemiluminescence matches the fluorescence spectrum of the monoanion of the starting hydrazide, (XII), under the reaction conditions. Equation (3) has been

proposed to account for this result.<sup>22</sup> This scheme requires production of excited phthalate and subsequent intermolecular energy transfer to the anion of the starting material from which emission occurs. The energy transfer apparently occurs between an excited phthalate ion hydrogen bonded to a phthalic hydrazide anion, since added fluorescers without acidic protons, such as acridone or the salt of *N*-methyl phthalic hydrazide, do not accept the excitation energy.

<sup>21</sup> J. Stauff and G. Hartmann, Ber. Bunsenges. Phys. Chem. 69, 145 (1965).

<sup>22</sup> E. H. White, D. F. Roswell, and O. C. Zafiriou, J. Org. Chem. 34, 2462 (1969).

$$(XII) \xrightarrow{\text{oxidation}} [\text{phthalate ion}] *$$

$$(XII) + (XII) \longrightarrow \text{phthalate ion} + (XII) *$$

$$(XII) + \dots + h\nu + (XII)$$

#### **Energy Transfer**

Energy transfer has also been shown to occur in the chemiluminescence of hydrazides<sup>23-26</sup> which are linked to some highly fluorescent groups. Compound (XIII) is a good example of this type of structure. In the case of

(XIII)

compound (XIII), oxidation of the hydrazide portion of the molecule (in protic or aprotic media) results in emission from the acridone moiety. This emission occurs because of rapid intramolecular singlet-singlet energy transfer from the electronically excited phthalate ion to the acridone portion of the molecule.

#### Reaction Mechanism

Information about the mechanism of oxidation of luminol and other hydrazides has been obtained in studies using both protic and aprotic media. In aprotic media (DMSO), the dinegative ion, (XIV), formed in a stepwise reaction with base is a crucial intermediate [Eq. (4)]. <sup>27</sup> Subsequent

- 23 E. H. White and D. F. Roswell, J. Am. Chem. Soc. 89, 3944 (1967).
- <sup>24</sup> E. H. White, D. R. Roberts, and D. F. Roswell, in "Molecular Luminescence" (E. C. Linn, ed.), 1st ed., p. 479. Benjamin, New York, 1969.
- <sup>25</sup> D. F. Roswell, V. Paul, and E. H. White, J. Am. Chem. Soc. 92, 4855 (1970).
- <sup>26</sup> D. R. Roberts and E. H. White, J. Am. Chem. Soc. 92, 4861 (1970).
- <sup>27</sup> E. H. White and M. M. Bursey, J. Am. Chem. Soc. 86, 941 (1964).

reaction of this intermediate with oxygen via some as yet undetermined pathway produces the phthalate ion in an excited state.

The reaction of the dinegative ion could follow several pathways, as indicated in Eqs. (5a)-(5d).

$$(XIV) + O_2 \longrightarrow R \longrightarrow N^{-} + O_2^{-}$$

$$(5a)$$

$$(XIV) + O_a \longrightarrow \bigvee_{R} \bigvee_{N} \bigvee_{N} (5b)$$

$$(XIV) + O_2 \longrightarrow \begin{array}{c} & & & \\ &$$

$$(XIV) \longrightarrow \begin{array}{c} O^{-} \\ N \\ N \end{array} \longrightarrow \begin{array}{c} O_{2} \\ N \\ N \end{array} \longrightarrow \begin{array}{c} O^{-} \\ N \\ N \end{array} \longrightarrow \begin{array}{c} (Sd) \\ (XVIII) \end{array}$$

The radical anion intermediate, (XV), in Eq. (5a) has also been proposed for the reaction in protic media when transition metal complexes are used as catalysts, and for the electrochemical oxidation. <sup>28</sup> In protic media it could be formed by the pathway shown in Eq. (6). <sup>7,29</sup> Alternatively, it has

been proposed<sup>30</sup> that the metal ion forms a complex with the hydrazide, which is then oxidized, and it has also been suggested that it is a  $HO_2^-M^{2+}$  complex which produces the hydrazide radical anion.<sup>31</sup>

The azaquinone proposed in Eq. (5b) has received considerable attention as a possible reaction intermediate; it was proposed as an intermediate in the first paper published on the chemiluminescence of luminol. Later workers, on the basis of kinetic evidence, have suggested azaquinones as intermediates in the persulfate/hydrogen peroxide-driven chemiluminescence of luminol as well as in the hypochlorite oxidation. Azaquinones have been trapped as their cyclopentadiene adducts, Azaquinones have been trapped as their cyclopentadiene adducts, agent normally exhibit

$$\bigcap_{R} \bigvee_{j=1}^{N} \bigvee_{j=1}^{N}$$

chemiluminescence. These adducts have been identified by thin-layer chromatography; the yield is low, accounting for only a small fraction of the starting hydrazide.

(XIX)

- 28 B. Epstein and T. Kuwana, Photochem. Photobiol. 4, 1157 (1965).
- <sup>29</sup> P. B. Shevlin and H. A. Neufeld, J. Org. Chem. 35, 2178 (1970).
- <sup>30</sup> A. K. Babko and N. M. Lukovskaya, Ukr. Khim. Zh. 30, 508 (1964).
- <sup>31</sup> T. G. Burdo and W. R. Seitz, Anal. Chem. 47, 1639 (1975).
- <sup>32</sup> M. M. Rauhut, A. M. Semsel, and B. G. Roberts, J. Org. Chem. 34, 2431 (1966).
- 33 W. R. Seitz, J. Phys. Chem. 79, 101 (1975).
- <sup>34</sup> Y. Omote, T. Miyake, and N. Sugiyama, Bull. Chem. Soc. Jpn. 40, 2446 (1967).

Diazaquinones are reactive but isolable compounds.<sup>35-37</sup> They chemiluminesce in the presence of alkaline hydrogen peroxide and, unlike the hydrazides, do not require peroxide-decomposing catalysts or oxidizing agents. Comparison of the emission spectra from the chemiluminescence of the hydrazide and the corresponding diazaquinone show that the emitter (the phthalate ion) is the same in the two cases. The diazaquinone has been detected by stop-flow spectroscopy during the hypochlorite-catalyzed oxidation of 2,3-naphthalic hydrazide.<sup>38</sup> However, its detection is a function of the hydrogen peroxide concentration. At high hydrogen peroxide concentrations it is not detectable. Rate studies are difficult to do and indicate that the steps prior to the production of the diazaquinone are rate determining. The few facts that are available support the hypothesis that azaquinones are intermediates in the light-producing oxidation of the hydrazides.

The intermediates proposed in Eqs. (5c) and (5d), instead of being formed directly, could also be thought of as arising from reaction of the azaquinone with hydroperoxide ion or superoxide ion [Eq. (7)]. Evidence

has been presented that superoxide may be involved in the reaction of luminol with base and oxygen in aqueous media, since superoxide dismutase quenches the chemiluminescent reaction.<sup>39</sup>

<sup>35</sup> K.-D. Gundermann, Angew. Chem. 80, 494 (1968).

<sup>&</sup>lt;sup>36</sup> E. H. White, E. G. Nash, D. R. Roberts, and O. C. Zafiriou, J. Am. Chem. Soc. 90, 5932 (1968).

<sup>&</sup>lt;sup>37</sup> K.-D. Gundermann, in "Chemiluminescence and Bioluminescence" (M. J. Cormier, D. M. Hercules, and J. Lee, eds.), p. 209. Plenum, New York, 1973.

<sup>&</sup>lt;sup>38</sup> C. Lee, unpublished results, 1974.

<sup>39</sup> E. K. Hodgson and I. Fridovich, Photochem. Photobiol. 18, 451 (1973).

Singlet oxygen, produced via dye sensitization, has been shown to cause luminol chemiluminescence in aqueous base.  $^{40-42}$  Singlet oxygen produced by laser irradiation produces only very weak chemiluminescence from luminol, and 3-aminophthalate is not the major product. It has been suggested that emission arising in the dye sensitization experiments is due to radicals being produced concurrently with singlet  $O_2$ .

The oxidation of hydrazides under mild conditions can produce products other than phthalate ion. Reaction of phthalic hydrazide with base and potassium ferricyanide or hydrogen peroxide-ammonium hydroxide-copper has been shown to produce phthalaldehydic acid, (XX), as well as phthalate ion itself.<sup>43</sup> Oxidation of the naphthalic hydrazide, (XXI), with

basic hypochlorite mixtures also produces the corresponding acid aldehyde.<sup>38</sup> The formation of the aldehyde is not surprising, since diacylimides are known to react with bases to produce one equivalent of acid and one of aldehyde [Eq. (8)].<sup>44</sup>

The yield of aldehyde in the hydrazide reactions decreases at the higher hydrogen peroxide concentrations that lead to enhanced light emission. Thus, the aldehyde may be a product of a dark reaction and the interception of species (XXII) may be directly on the light-emission pathway [Eq. (8)].

(XIV) 
$$\begin{array}{c} O \\ O \\ C \\ C \\ O \end{array}$$

$$\begin{array}{c} O \\ O \\ C \\ C \\ O \end{array}$$

$$\begin{array}{c} O \\ O \\ C \\ O \\ \end{array}$$

$$\begin{array}{c} O \\ O \\ C \\ O \\ \end{array}$$

$$\begin{array}{c} O \\ O \\ O \\$$

- 40 K. Kuschnir and T. Kuwana, Chem. Commun. 1969, 193 (1969).
- <sup>41</sup> I. B. C. Matheson and J. Lee, Photochem. Photobiol. 12, 9 (1970).
- <sup>42</sup> I. B. C. Matheson and J. Lee, *Photochem. Photobiol.* 24, 605 (1976).
- <sup>43</sup> T. Huang, unpublished results, 1972.
- 44 C. Niemann and J. T. Hays, J. Am. Chem. Soc. 74, 5796 (1952).

In summary, it appears that dianion (XIV) is involved in aprotic oxidations, the radical anion (XV) in protic ones, and the azaquinones very probably are involved in the light pathway.

#### Monoacylhydrazides

Noncyclic nonoacylhydrazides, such as compounds (XXIII)-(XXV), have been reported to be chemiluminescent. 45,46 The oxidizing systems used are the same as those used for the cyclic hydrazides. The

monoacylhydrazides are moderately chemiluminescent compounds; compounds (XXIV) and (XXV) have  $\Phi_{\text{Cl}} = 0.003$  (in comparison, the value for luninol is 0.01).

The pathway leading to light emission appears to be different from that of luminol and the cyclic hydrazides, <sup>47</sup> and thus these compounds are not good models for the luminol chemiluminescence. The emitter in the case of compound (XXV) corresponds to the fluorescence of acridone and its anion. Thus the most efficient light-producing reaction requires a decarbonylation or decarboxylation. One crucial intermediate would be the acyl anion (XXVI), which could lose carbon monoxide and react with oxygen to

45 E. H. White, M. M. Bursey, D. F. Roswell, and J. H. M. Hill, J. Org. Chem. 32, 1198 (1967).

<sup>46</sup> E. Rapaport, M. W. Cass, and E. H. White, J. Am. Chem. Soc. 94, 3153 (1972).
 <sup>47</sup> In an earlier report <sup>46</sup> it was erroneously proposed that the mechanism of oxidation of (XXIV) and (XXV) were the same as for the cyclic hydrazides, and that the corresponding carboxylate ion was the light emitter.

form the excited state of acridone anion<sup>48,49</sup> [(XXVII), Eq. (9)]. Alternatively, the acyl anion could react with oxygen and then decarboxylate via a dioxetane intermediate with formation of an excited state<sup>50-52</sup> [Eq. (10)]. Note that acridine carboxylic acid is only weakly fluorescent and that it may be formed via the luminol pathway also, but the low quantum yield to be expected precludes detection of this pathway.<sup>47</sup>

(XXVII) 
$$O_2$$
 $O_2$ 
 $O_3$ 
 $O_4$ 
 $O_5$ 
 $O_7$ 
 $O_8$ 
 $O_9$ 
 $O_9$ 
 $O_9$ 
 $O_9$ 

$$(XXVI) \xrightarrow{O_2} O \xrightarrow{O} O \xrightarrow{O} O \xrightarrow{N} (XXVII)^*$$

$$(10)$$

The decarbonylation (decarboxylation) pathway does not appear to be operative in the case of the cyclic hydrazides, since naphthalic hydrazide, (XXI), yields naphthalate excited states, but not those of 3-hydroxyl-2-naphthoic acid.

<sup>&</sup>lt;sup>48</sup> G. A. Russell and E. G. Janzen, J. Am. Chem. Soc. 84, 4153 (1962).

<sup>&</sup>lt;sup>49</sup> G. A. Russell and A. G. Bemis, J. Am. Chem. Soc. 88, 5491 (1966).

<sup>50</sup> F. McCapra, Acc. Chem. Res. 9, 201 (1976).

<sup>&</sup>lt;sup>51</sup> N. J. Turro, P. Lechtken, N. E. Schore, G. Schuster, H. C. Steinmetzer, and A. Yekta, Acc. Chem. Res. 7, 97 (1974).

<sup>52</sup> E. H. White, J. D. Miano, C. J. Watkins, and E. Breaux, Angew. Chem., Int. Ed. 13, 229 (1974).